# Structural Brain Changes in Pre-Clinical FTD *MAPT* Mutation Carriers

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- 14 Abstract.
- Background: Frontotemporal dementia (FTD) is the second most common cause of early-onset neurodegenerative dementia.
- Several studies have focused on early imaging changes in FTD patients, but once subjects meet full criteria for the FTD diagnosis, structural changes are generally widespread.
- 18 **Objective:** This study aims to determine the earliest structural brain changes in asymptomatic *MAPT* MUTATION carriers.
- <sup>19</sup> Methods: This is a cross-sectional multicenter study comparing global and regional brain volume and white matter integrity
- in a group of *MAPT* mutation preclinical carriers and controls. Participants belong to multiple generations of six families
- with five *MAPT* mutations. All participants underwent a medical examination, neuropsychological tests, genetic analysis,
   and a magnetic resonance scan (3T, scout, T1-weighted image followed by EPI (BOLD), MPRAGE, DTI, FLAIR, and ASL
- 23 sequences).
- Results: Volumes of five cortical and subcortical areas were strongly correlated with mutation status: temporal lobe (left
- amygdala, left temporal pole), cingulate cortex (left rostral anterior cingulate gyrus, right posterior cingulate), and the lingual
- gyrus in the occipital lobe. We did not find significant differences in whole brain volume, white matter hyperintensities
- volume, and white matter integrity using DTI analysis.
- 28 **Conclusion:** Temporal lobe, cingulate cortex and the lingual gyrus seem to be early targets of the disease and may serve as
- <sup>29</sup> biomarkers for FTD prior to overt symptom onset.
- 30 Keywords: Atrophy, brain atrophy, early detection, frontotemporal lobar degeneration, *MAPT* mutation

# 31 INTRODUCTION

Frontotemporal dementia (FTD) is the second most common cause of early-onset neurodegenerative dementia [1]. Up to 40% of FTD cases are associated with an autosomal dominant pattern of inheritance. Mutations in over eight genes have been identified in FTD, including progranulin (*GRN*), chromosome 9 open reading frame (*C9orf72*), and microtubule-associated protein tau (*MAPT*) genes [2, 3]. Neuroimaging has been explored as a potential biomarker to identify patients in initial phases of

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neurodegenerative diseases and to measure biologi-42 cal change over time [4]. Several studies have focused 43 on early imaging changes in FTD patients [5-10], 44 but once subjects meet full criteria for FTD diagno-45 sis, structural changes are generally widespread [11]. 46 Studying mutation carriers who are asymptomatic 47 or transitioning from asymptomatic to symptomatic 48 (what we will refer to as "preclinical") [12] may 49 allow for characterization of the earliest neuroimag-50 ing changes in disease [13-17]. 51

Studies specifically addressing preclinical neu-52 roimaging features in MAPT mutation carriers are 53 scarce. Some studies [18, 19] have demonstrated 54 early insular atrophy, and others early medial tem-55 poral degeneration [20, 21]. Also, some studies have 56 reported early white matter changes in preclinical 57 FTD patients [22-25], but those performed specifi-58 cally in MAPT mutation carriers have not [26]. Our 50 institution has access to a relatively large, broadly 60 phenotyped group of MAPT mutation carriers and 61 familial matched controls that brings the opportunity 62 to study early imaging changes in a well char-63 acterized population to address the inconsistencies 64 found in previous studies. With that purpose, we 65 compare cortical and subcortical gray matter vol-66 umes as well as white matter hyperintensities and 67 tract-integrity between MAPT mutation carriers and 68 demographically-matched familial controls. 69

# 70 MATERIALS AND METHODS

#### 71 Participants

Participants were recruited from an active, longitu-72 dinal research protocol (R01NS076837) that includes 73 multiple generations of six families with MAPT muta-74 tions: V337M (c.2014G>A), P301L (c.1907C>T), 75 Exon 10+14 C>T, Exon 10+15 C>T, and Exon 76 10+16 C>T. Members of these families live 77 throughout areas of the United States and Europe. 78 Those who consented to participate in the study were 79 followed at Columbia University Medical Center, the 80 University of Michigan, and the Dublin Neurological 81 Institute. 82

Sixty subjects were enrolled and 56 completed
the baseline visit, during which genetic, biological,
neuroimaging, and clinical data were collected. Four
subjects did not complete the study due to rejection
to undergo all the exams. No imaging studies were
rejected for the analysis due to quality issues. Sample
characteristics are displayed in Table 1.

#### Clinical assessments

Most participants, and investigators whenever possible, were blind to carrier status. A full history and physical and neurological examination was performed by one of the study physicians at the enrolling clinical site. The Clinical Dementia Rating (CDR) Scale, including the language and behavior components (CDR<sup>®</sup> Plus NACC FTLD), as well as cognitive, behavioral, and psychiatric measures was completed as part of this evaluation (Table 1). Informants provided input for the clinical assessment and participated in the completion of behavioral interviews administered by the study coordinator. Participant cognitive status was characterized using tests from the National Alzheimer's Coordinating Center (NACC) UDS 2.0 Neuropsychological Battery and NACC UDS 2.0 FTLD Module. This evaluation included assessment of memory function (Mini-Mental State Examination, Selective Reminding Test immediate and delayed recall, Selective Reminding Test discriminability index), verbal function (categorical fluency, Boston Naming Test, Controlled Oral Word Association (COWA)), visual cognition (Benson figure), executive function (Trail A, Trail B, COWA, 20 Q's, Design Fluency, Graphic Pattern Generation), social abilities (Social Norms Questionnaire 22, Empathic Concern Score, Perspective Taking Score, Revised Self-Monitoring Scale), other frontal lobe functional tests (Remote Associates Test) as well as depression and anxiety (Neuropsychiatric Inventory).

## Genetic analysis

Blood was collected through standard phlebotomy procedures at the Columbia University Medical Center (Irving Center for Clinical and Translational Research), University of Michigan, and the Dublin Neurological Institute. Fifteen cc of blood were collected, including citrate tubes for DNA isolation and heparin tubes for plasma isolation. DNA was prepared from whole blood using standard protocols in the Columbia Human Genetics Resources Core. Polymerase chain reaction (PCR) and amplification were performed in all samples. The PCR and sequencing primers used for amplification and sequencing were designed using the software Primer 3 (http://frodo.wi.mit.edu/primer3/). Cycle sequencing in forward and reverse directions was performed on purified PCR products and run on an ABI 3730 genetic analyzer (Applied Biosystems,

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Demographic information about age, gender, and education. Results of neuropsychological tests adjusted (by age, gender, and education). By Fisher exact test/2-sided Wilcoxon test, there is not significant difference of age, gender, and education between carriers and non-carriers

Clinical features	Non-carrier $(n=44)$		Carrier $(n=12)$		Carrier (CDR 0) (n=6)		Carrier (CDR 0.5) (n=6)	
Gender	Count	%	Count	%	Count	%	Count	%
Female	24	54.55	8	66.67	4	66.67	4	66.67
Male	20	45.45	4	33.33	2	33.33	2	33.33
	Mean	SD	Mean	SD	Mean	SD	Mean	SD
Age	44.64	13.40	48.83	13.37	39.00	11.52	58.67	5.32
Education	15.18	2.33	15.25	2.14	15.33	2.07	15.17	2.40
Neuropsychiatric tests	Mean	SD	Mean	SD	Mean	SD	Mean	SD
SNQ22	-0.29	0.94	-1.18	1.3	-1.12	1.77	-1.24	0.74
EC	-0.02	1.1	-0.45	1.12	0.17	1.3	-1.07	0.39
PT	0.08	1.09	-1.06	1.27	-0.4	1.42	-1.72	0.7
RSMS (Z-score)	-1.6	1.2	-2.67	1.94	-1.49	1.93	-3.86	1.11
SRT <sup>1</sup>	46.06	10.44	32.47	19.87	50.42	4.34	14.52	8.73
SRT <sup>2</sup>	48.33	8.81	37.62	18.22	53.26	6.16	21.98	10.26
MMSE	0.59	0.73	-0.25	1.45	0.25	1.13	-0.75	1.66
Cat. F (Animals)	-0.46	0.79	-1.32	1.27	-0.35	0.54	-2.28	1.01
Trail-A (Z-score)	-0.35	0.61	-1.09	1.41	-0.48	0.44	-1.69	1.82
Trail-B (Z-score)	-0.4	0.74	-0.89	1.01	-0.56	0.3	-1.23	1.37
BNT (Z-score)	-0.53	0.67	-2.81	2.67	-0.73	0.71	-4.88	2.2
Benson Figure Copy (Z-score)	0.73	1.03	0.31	1.2	0.59	1.05	0.03	1.37
CFL	47.2	9.32	42.25	12.06	47	9.36	37.5	13.35
SRT	0.97	0.04	0.87	0.19	0.99	0.03	0.75	0.21
20 Q's	11.69	2.48	10.17	3.33	12	1.79	8.33	3.61
RAT	15.98	0.15	15.17	1.34	15.83	0.41	14.5	1.64
DF	17.41	1.88	13.82	7.61	18.67	1.51	8	8.03
GPG <sup>1</sup>	6.83	4.07	3.7	4.6	3.7	4.6	3.7	5.14
GPG <sup>2</sup>	309.2	181.1	310.9	206.7	261.6	155.9	360.2	256.4
NPI*	Non-carriers		Carriers		Carriers CDR 0		Carriers CDR 0.5	

NPI*	Non-carriers	Carriers	Carriers CDR 0	Carriers CDR 0.5
	(n = 44)	( <i>n</i> = 12)	(n = 6)	(n = 6)
Agitation	4.88%	45.45%	0.00%	83.33%
Anxiety	9.76%	40.00%	20.00%	60.00%
Apathy	2.44%	36.36%	0.00%	66.67%
Appetite	9.76%	27.27%	0.00%	50.00%
Delusions	2.44%	0.00%	0.00%	0.00%
Depression/dysphoria	9.76%	27.27%	20.00%	33.33%
Disinhibition	4.88%	45.45%	0.00%	83.33%
Elation	0.00%	27.27%	0.00%	50.00%
Hallucinations	0.00%	9.09%	0.00%	16.67%
Irritability	4.88%	36.36%	0.00%	66.67%
Motor disturbance	2.44%	18.18%	0.00%	33.33%
Nighttime behaviors	4.88%	18.18%	0.00%	33.33%

SNQ22, Social Norms Questionnaire (Z-score); EC, Interpersonal Reactivity Index Empathic Concern Score (Z-score); PT, Interpersonal Reactivity Index Perspective Taking Score (Z-score); RSMS, Revised Self-Monitoring Scale (Z-score); SRT<sup>1</sup>, Immediate Recall T-score (total score); SRT<sup>2</sup>, Delayed Recall T-score (Total score); MMSE, Mini-Mental State Examination Total score (Z-score); Cat.F Animals, Category Fluency for Animals (Z-score); Trail-A, Trail Making Test part A (Z-score); Trail-B, Trail Making Test part B (Z-score); BNT, Boston Naming Test (Z-score); Benson Figure (Z-score); CFL, Controlled Oral Word Association (COWA) Test using C, F, and L; SRT, Selective Reminding Test Discriminability index; 20 Q's, 20 questions from DKEFS Total Weighted Achievement - Scaled Score; RAT, Remote Associates Test Total Score; DF, Design Fluency total correct score; GPG<sup>1</sup>, Graphic Pattern Generation Perseveration Distance; GPG<sup>2</sup>, Graphic Pattern Generation Perseveration Time (in seconds), NPI, Neuropsychiatric Inventory. \*3 non-carriers with CDR 0 (ES012, ES024, ES036) and 1 carrier with CDR 0 (ES038) do not have NPI-Q data.

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Foster City, CA). Sequence chromatograms were viewed and genotypes determined using Sequencher (Genecodes). Samples were stored frozen at -80°C.
Subjects did not receive DNA results as part of the current study. Patients were screened for mutations in other genes associated with FTD besides *MAPT*.

# Imaging procedures

A number of imaging modalities were acquired during a 1 h magnetic resonance (MR) scan performed in NY and Dublin in a 3.0T Philips Achieva Quasar Dual Magnet using a 240 mm field of view.

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A scout, T1-weighted image was first acquired to 149 determine patient position, followed by echo pla-150 nar imaging (blood oxygenation level dependent), 151 magnetization-prepared rapid acquisition with gra-152 dient echo (MPRAGE), diffusion tensor imaging 153 (DTI), fluid-attenuated inversion recovery (FLAIR), 154 and arterial spin labeling sequences. The following 155 parameters were used: 156

- \* T1-weighted: repetition time = 20 ms, echo time = 2.1 ms, field of view = 240 cm, and  $256 \pm 160$  matrix with 1.3 mm slice thickness.
- \* FLAIR: repetition time = 11,000 ms, echo 160 time = 144.0 ms, inversion time = 2800 ms, field 161 of view = 25 cm, 2 NEX, and  $256 \pm 192$  matrix 162 with 3 mm slice thickness. 163
  - \* DTI: repetition  $time = 11032 \, ms.$ echo time = 69 ms, acquisition time 6 mins, slice thickness 2 mm.

A neuroradiologist reviewed each subject's MRI 167 scan for clinical abnormalities. Scanning proce-168 dures were standardized between all centers using 169 methods previously described [27] conducted in 170 person by a radiologist from CUMC. Structural 171 imaging measures of global and regional brain 172 volume were derived from each individual's T1-173 weighted MPRAGE image using Freesurfer software 174 (http://surfer.nmr.mgh.harvard.edu/). For brain vol-175 ume calculations, we used the procedures of Walhovd 176 et al. [28] to automatically assign a neuroanatomical 177 label to each voxel, with results comparable to man-178 ual labeling. From this labeling, volumetric regions 179 of interest (ROIs) were defined. The calculated vol-180 ume within each region was adjusted for variations 181 in individual global brain volume with a measure 182 of total intracranial volume (TIV). We decided to 183 compare values of all the different areas, without 184 defining ROIs *a priori*. This method allows an unbi-185 ased assessment of patterns of atrophy across the 186 whole brain without limiting the number of potential 187 measurements performed in the study [29]. In order 188 to measure white matter hyperintensities, each par-189 ticipant's FLAIR image was skull stripped and after, 190 voxel intensity values of the remaining image were 191 analyzed using a Gaussian curve. Hyper-intense vox-192 els were defined using a threshold of 2.1 SD above the 193 mean intensity. They were labeled and measured in 194 cubic centimeters multiplying the number of voxels 195 for the voxel's dimensions. 196

DTI data was processed in FMRIB's Diffusion 197 Toolbox (FDT), distributed as part of FMRIB's 198 Software Library [30], by first preprocessing with 199

eddy-current correction followed by fitting of the 200 DTI model to the preprocessed data. To align all 201 subjects into the same common space, tract-based 202 spatial statistics [31] was run on the fractional 203 anisotropy (FA) maps using the nonlinear registra-204 tion tool FNIRT [32, 33] and then the mean FA image 205 was created and thinned to create a mean FA skeleton, 206 representing the centers of all tracts common to the 207 group. Each subject's aligned FA data was then pro-208 jected onto this skeleton and created a skeletonized map per subject. To extract 20 tracts of interest, Johns Hopkins University (JHU) white matter tractography 211 atlas [34] was used as masks to obtain the mean FA for each tract for each participant.

#### Standard protocol approvals, registrations, and participant consents

Approval for this study was obtained from the appropriate IRB and ethics boards of Columbia and University of Michigan Medical Centers and the Dublin Neurological Institute. Written informed consent was obtained from all participants.

### Statistical analysis

For statistical analysis, stats (v 3.3.0), glmnet (v 2.5.0) [35], and pROC (v 1.9.1) [36] packages implemented in R (v.3.3.1) were used. As a primary analysis, we hypothesized group differences in 16 bilateral subcortical, 68 bilateral cortical ROI volumes, and 5 white matter hyperintensity volumes (frontal, temporal, parietal, occipital, and basal ganglia). The volumetric measures were corrected for age and TIV. Group comparison between mutation carriers and non-carriers was conducted using Wilcoxon test followed by multiple comparison correction controlling for false discovery rate [37]. Wilcoxon test analysis followed by multiple comparison was also performed for each ROI for both CDR 0 and CDR 0.5 carriers, residualized for age and TIV. Regarding DTI results, group analysis was conducted on the mean FA for the 20 tracts to compare carriers versus noncarriers. Age was included as a covariate to remove the confound of age and correction for multiple comparisons was performed using the false discovery rate [37].

We also evaluated discriminability of the volumes using penalized logistic regression with elastic-net [38] and receiver operating characteristic (ROC) analysis, adjusted by clinical covariates. To select tuning parameters for elastic-net, leave-one-out cross

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validation was used ( $\alpha = 1$ ,  $\lambda = 0.0469$ ). We did a 500 iteration bootstrapping to calculate the consistency of selecting each of the ROIs into the multivariate model.

#### 252 **RESULTS**

We analyzed data from 56 participants belonging 253 to five families carrying MAPT mutations. Twelve 254 subjects were determined to be carriers of the fol-255 lowing mutations: P301L (one subject), Exon 10+16 256 C>T (one subject), Exon 10+15 C>T (two sub-257 jects), V337M (c.2014G>A) (four subjects), and 258 Exon 10 + 14 C > T (four subjects). Forty-four sub-259 jects were non-carriers. Demographic characteristics 260 of the sample and performance in neuropsycholog-261 ical testing are displayed in Table 1. Subjects were 262 considered preclinical if they did not fulfill FTD diag-263 nostic criteria. We use the term "preclinical" rather 264 than "presymptomatic" as this group includes those 265 that scored 0 and 0.5 in CDR. CDR = 0 carriers had no 266 cognitive or behavioral impairment, but all CDR = 0.5267 carriers had one or more abnormal neuropsycholog-268 ical score, and most also had questionable or mild 269 behavioral impairment as indicated on the Behav-270 ior, Comportment, and Personality rating of the CDR 271 Supplemental scores (FTLD-CDR) [39]. However, as 272 these abnormalities were considered as questionable 273 by raters and they did not fulfill diagnostic criteria for 274 FTD, patients with CDR score of 0.5 were included 275 in the preclinical group. There were no significant 276 differences in age, gender, and years of education 277 between the groups. 278

For each ROI, Freesurfer-derived raw volumes 279 were converted to percentage of TIV prior to the 280 analyses. The resulting data were analyzed using 281 two models, a Wilcoxon analysis and a multivari-282 ate elastic-net model. In Wilcoxon analysis, 32 of 283 89 measures show significant between-group dif-284 ferences after multiplicity adjustment (Table 2), 285 including caudate, putamen, hippocampus, amyg-286 dala, nucleus accumbens, and several regions of 287 the frontal and temporal lobes. Over 500 itera-288 tion bootstrapping, five measures were constantly 289 selected (with over 80% selection rate), demonstrat-290 ing a potential strong association with mutation status 291 (Table 2, Fig. 1). These five ROIs are left amygdala 292 (selection percentage 0.83), rostral anterior cingu-293 late (selection percentage 0.91), posterior cingulate 294 (selection percentage 0.81), temporal pole (selection 295 percentage 0.92), and lingual volume (selection per-296

centage 0.83). Among these five, the lingual area and posterior cingulate did not come out significant in Wilcoxon's test, likely because they are only strong predictors of mutation status when other ROIs are controlled. No significant difference was found in whole brain volume between mutation carriers and non-carriers. Results remained the same after recalculating bilateral selection percentage over 500 bootstrap iterations as when considering both hemispheres separately. Therefore, for those ROIs, the left and right sides were not competing. When considering the sum of ROIs on both sides, four measures were selected over 80% selection rate: white matter hyperintensities in the occipital lobe (selection percentage 0.862), temporal pole volume (selection percentage 0.97), lingual volume (selection percentage 0.876). and posterior cingulate volume (selection percentage 0.84).

We performed a subgroup analysis comparing CDR 0 and CDR 0.5 carriers (2-sided Wilcoxon Test after adjustment for multiple comparisons). We found significant differences in 18/89 measures after multiplicity adjustment with larger volumes in CDR 0 subgroup (shown in Table 3 with an \*, values available in the Supplementary Table 1). Most of these ROIs are the same that were significantly different between carriers and non-carriers. Only left superior frontal gyrus (CDR0=1.26 versus CDR0.5=0.97, p = 0.04) and right precentral gyrus (CDR0 = 0.77) versus CDR0.5 = 0.63, p = 0.049) showed differences between CDR subgroups that were not present when comparing carriers and non-carriers. These results should be considered with caution, as the number of subjects for the subgroup analysis is very low.

We did not find significant differences between carriers and non-carriers in the volume of white matter hyperintensities. There was only a weak difference after adjusting results by age and TIV, which did not survive after adjustment for multiple comparisons and elastic-net analysis (Table 2). We found a significant difference for white matter hyperintensities in the basal ganglia between CDR 0 and CDR 0.5 preclinical carriers (CDR 0=2.69E-05 versus CDR 0.5=4.49E-06; p=0.03) after adjustment for multiple comparisons.

DTI measures were available for 9 carriers and 40 controls due to technical issues during the neuroimaging exam. After comparing DTI results between groups, we found significant differences between carriers and non-carriers for the left cingulum at the cingulate gyrus ( $0.579 \pm 0.055$  versus  $0.609 \pm 0.034$ ; p = 0.041), right cingulum at the cingulate gyrus

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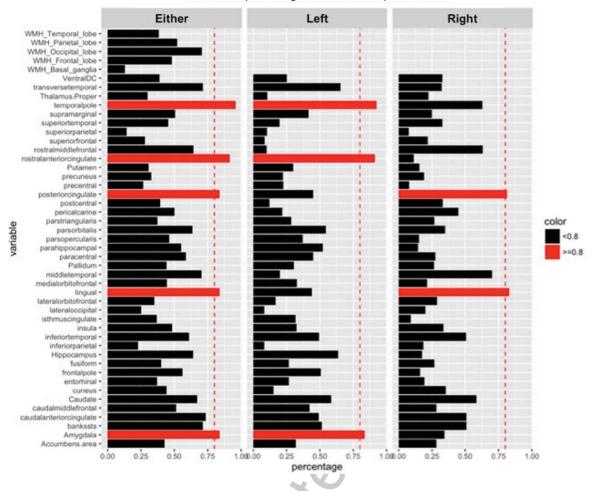
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#### Table 2

Wilcoxon analysis and multivariate Elastic-net analysis. In Wilcoxon analysis 32/89 measures show significant between group differences after multiplicity adjustment. In Elastic-net model, 10/89 ROIs are selected into the model. Over 500 iteration bootstrapping, 5 measures are constantly selected (with over 80% selection rate) demonstrating a potential strong association with mutation status. The column 'Either' is a surrogate measure of the % selection for either left or right hemisphere of the specific ROI. When considering bilateral selection %, results remain the same as considering both hemispheres separately

Region		2-sided Wilcoxon test results			Percentage of selection			
White matter	Raw	Raw data		multiple adjustment:		over 500 Bootstrap		
hyperintensities		ituw uuu		FDR (HL:<0.05)		iterations		
Frontal_lobe	0.4	0.4129		· · · · · ·				
Parietal_lobe		0.4129 0.1909		0.4821 0.2879		0.480 0.522		
Occipital_lobe				0.2467				
Temporal_lobe		0.0634		0.0794	0.706 0.384			
1		0.0515 0.1494				0.384 0.130		
WMH_Basal_ganglia*			0.2770 <sup>1</sup>		Laft	Right		
Gray matter	Left	Right	Left	Right 0.4349	Left 0.106		Either 0.298	
Thalamus.	0.1736	0.2151	0.1665			0.222		
Caudate	0.0234	0.0396	0.0166	0.0223	0.584	0.584	0.670	
Putamen	0.0246	0.0197	0.0243	0.0364	0.298	0.156	0.306	
Pallidum	0.4834	0.3081	0.7957	0.6757	0.304	0.264	0.442	
Hippocampus	0.0040	0.0186	0.0281 <sup>1</sup>	0.02631	0.636	0.174	0.640	
Amygdala	0.0007	0.0137	0.0125 <sup>1</sup>	0.03131	0.834	0.342	0.838	
Accumbens area	0.0074	0.0040	0.0405	0.02411	0.320	0.282	0.426	
Ventral diencephalon	0.4236	0.4834	0.6564	0.7736	0.252	0.326	0.390	
Banks of superior temporal sulcus	0.0918	0.0208	0.2233	0.0271	0.514	0.510	0.714	
Caudal ant.Cingulate <sup>†</sup>	0.0999	0.4470	0.0815	0.4957	0.490	0.510	0.736	
Caudal middle frontal	0.4470	0.0233	0.7605	0.0736	0.422	0.282	0.514	
Cuneus	0.0770	0.1317	0.1362	0.2664	0.152	0.352	0.442	
Entorhinal areal	0.0057	0.1228	0.0192	0.2326	0.266	0.192	0.372	
Fusiform gyrus	0.0737	0.0274	0.0794	0.0271	0.268	0.266	0.402	
Inf. parietal gyrus <sup>†</sup>	0.0558	0.1777	0.0949	0.2233	0.082	0.184	0.226	
Inf. temporal gyrus	0.0042	0.0030	$0.0125^{1}$	0.0159 <sup>1</sup>	0.494	0.506	0.610	
Isthmus cingulate	0.4122	0.5425	0.5165	0.9399	0.316	0.090	0.370	
Lateral occipital gyrus	0.2987	0.4122	0.1360	0.4889	0.082	0.200	0.252	
Lateral orbitofrontal	0.0222	0.0131	0.0488	0.0166 <sup>1</sup>	0.168	0.286	0.348	
Lingual gyrus	0.6012	0.9443	0.9399	0.5813	0.440	0.828	0.838	
Medial orbitofrontal	0.0305	0.0427	0.0488	0.0736	0.326	0.212	0.444	
Middle temporal	0.0050	0.0030	0.0281	0.01251	0.200	0.700	0.702	
Parahippocampal gyrus	0.0515	0.2677	0.0488	0.4089	0.522	0.146	0.552	
Paracentral gyrus	0.1438	0.2805	0.2230	0.5165	0.448	0.274	0.586	
Pars opercularis	0.1291	0.1438	0.1932	0.2099	0.372	0.152	0.460	
Pars orbitalis	0.0103	0.0504	0.0166	0.1299	0.544	0.348	0.636	
Pars triangularis	0.2120	0.0918	0.2716	0.2879	0.284	0.266	0.374	
Pericalcarine gyrus	0.2463	0.3229	0.6757	0.5165	0.216	0.446	0.500	
Postcentral gyrus	0.5605	0.3229	0.6634	0.5884	0.122	0.330	0.394	
Post. cingulate gyrus <sup>¶</sup>	0.7007	0.8981	0.9764	0.9294	0.448	0.814	0.838	
Precentral gyrus	0.3899	0.1317	0.4089	$0.1362^{1}$	0.224	0.078	0.266	
Precuneus	0.3683	0.2084	0.2517	0.2279	0.222	0.190	0.326	
Rost. ant.cing. gyrus <sup>#</sup>	0.0022	0.1170	0.00421	0.2056	0.912	0.170	0.920	
Rostral mid.frontal gyrus**	0.0879	0.0305	0.0405	0.2050 0.0159 <sup>1</sup>	0.100	0.630	0.642	
Superior frontal gyrus	0.0619	0.0613	0.0837 <sup>1</sup>	0.0562	0.086	0.030	0.278	
Superior parietal gyrus	0.0019	0.0013	0.0857	0.0362	0.080	0.218	0.142	
Sup. temporal gyrus <sup>††</sup>	0.0102	0.0068	0.0159	0.0141 <sup>1</sup>	0.196	0.328	0.454	
Supramarginal gyrus	0.4352	0.0131	0.6757	0.0488	0.414	0.250	0.506	
Frontal pole	0.7494	0.2048	0.6757	0.4089	0.506	0.160	0.560	
Temporal pole	0.0018	0.0195	0.0159	0.0313 <sup>1</sup>	0.924	0.628	0.958	
Transverse temporal gyrus	0.0704	0.4236	0.1360	0.7736	0.654	0.322	0.712	
Insula	0.0260	0.0208	0.0159 <sup>1</sup>	0.0488 <sup>1</sup>	0.324	0.334	0.484	

The 5 ROIs are: left amygdala, left rostral anterior cingulate, left temporal pole, right lingual gyrus, andright posterior cingulate gyrus. Among these 5, right lingual gyrus and right posterior cingulate did not come out significant in Wilcoxon's test, probably because they are only a strong predictor of the mutation status when other ROIs are controlled. \*White matter hyperintensities basal ganglia; <sup>†</sup>caudal anterior cingulate; <sup>†</sup>Inferior parietal gyrus; Inferior temporal gyrus; <sup>¶</sup>Posterior cingulate gyrus; <sup>#</sup>Rostral anterior cingulate gyrus; <sup>†\*</sup>Rostral middle frontal gyrus; <sup>††</sup>Superior temporal gyrus. <sup>1</sup>ROIs that showed significant differences between CDR 0 carriers and CDR 0.5 carriers.



#### Selection percentage in 500 bootstrap iterations

Fig. 1. Multivariate analysis: Elastic net. Percentage of selection over 500 Bootstrap iterations. In Elastic-net model, 10/89 ROIs are selected into the model. Over 500 iteration bootstrapping, 5 measures are constantly selected (with over 80% selection rate), demonstrating a potential strong association with mutation status. This is a recalculation of selection percentage based on the bootstrap results. The column 'Either' is a surrogate measure of the % selection for either left or right hemisphere of the specific ROI. When considering bilateral selection %, the results remain the same as considering both hemispheres separately. It could be shown from the graph that for those ROIs, the bilateral ROIs seems not competing with each other.

 $(0.531 \pm 0.053 \text{ versus } 0.562 \pm 0.038; p = 0.045)$ , and left cingulum at the hippocampus  $(0.522 \pm 0.074 \text{ versus } 0.563 \pm 0.047; p = 0.039)$ . However, after adjusting these results for age, the differences were no longer significant.

# 354 DISCUSSION

In this study, we report early structural changes in pre-clinical *MAPT* mutation carriers measured using voxel-based morphometry and DTI analysis. Prior to overt symptom onset, *MAPT* mutation carriers in our sample showed volume differences in almost 30% of the 89 regions explored, including basal ganglia (caudate, putamen), temporal lobe (in particular medial temporal lobe), and some areas of the cingulate gyrus and the medial frontal lobe. Most areas were equally affected in both hemispheres, with a symmetrical distribution that has been previously described in *MAPT* mutation patients [19, 40]. Using a much more restrictive statistical analysis five regions were consistently associated with mutation status including the left temporal lobe (left amygdala, left temporal pole), bilateral cingulate cortex (left rostral anterior cingulate gyrus, right posterior cingulate), and the lingual gyrus in the occipital lobe.

Until now, studies regarding early structural changes in asymptomatic *MAPT* carriers have shown

inconsistent results: while several small case series 375 had reported neuroimaging differences in asymp-376 tomatic subjects, other studies did not find any 377 structural change [40, 41]. Those studies that found 378 differences point to early degeneration in the tempo-379 ral lobe, medial frontal lobe, and cingulate cortex. An 380 earlier study, published in 2010 by Miyoshi et al. [21], 381 found the medial temporal lobe and cingulate gyrus 382 affected (hippocampal atrophy and striatal dopamin-383 ergic dysfunction). The most recent work shows 384 also emerging grey matter temporal lobe changes 385 in MAPT asymptomatic mutation carriers [42]. Our 386 results agree partially with these findings as well as 387 with the largest study in asymptomatic mutation car-388 riers to date [19, 20], which found differences in 389 hippocampal and amygdala volumes as early as 15 390 years prior to expected symptom onset, in the tempo-391 ral lobe 10 years before expected onset and in insula 5 392 years before expected symptom onset. However, after 393 correction for multiple comparisons, only changes in 394 the insular area remained significant. Ours study sup-395 ports these previous results, confirming differences in 396 hippocampus, amygdala, the insular area, and tem-397 poral lobe between carriers and non-carriers, which 398 survived correction for multiple comparisons. More-399 over, we found significant differences between basal 400 ganglia volume (caudate, putamen), some areas of 401 the frontal lobe, and the cingulate gyrus. Our results 402 survived multiple comparison correction even though 403 we applied a much more restrictive statistical method 404 in order to determine the strongest association with 405 mutation status. If we consider only the elastic-406 net model results, the amygdala and temporal lobe 407 remain affected in our sample, whereas there were 408 no differences for hippocampus and insula volumes. 409 Considering that the insula was the last structure 410 affected in the GENFI study, it is possible that the 411 carriers in our cohort had not yet reached that stage of 412 progression. Both studies have important differences 413 regarding sample characteristics, statistical method-414 ology, and imaging analysis. Our sample includes 415 fewer asymptomatic carriers than the GENFI study; 416 however, subjects came from only five families car-417 rying five mutations, as opposed to the GENFI 17 418 different families of MAPT carriers carrying ten dif-419 ferent mutations. Considering that FTD is a protean 420 pathology [11] in which disease phenotypes may 421 vary substantially between families and individuals 422 with different mutations [43], our sample's charac-423 teristics can add to the body of knowledge currently 424 available. Regarding neuroimaging, our study was 425 performed at only three sites and we used only one 426

type of MRI scan, which increases the consistency 427 of the results. We examined the volume not only 428 of the brain lobes as a whole, but also of specific 429 areas in each lobe. Regarding white matter changes, 430 our results agree with the findings of the GENFI 431 cohort [26], which did not report any difference in 432 white matter hyperintensities either in clinical and 433 preclinical MAPT mutation carriers when compared 434 to controls. Interestingly, our analysis was performed 435 using FLAIR sequence, which is the standard for the 436 study of white matter hyperintensities and is usu-437 ally available in clinical practice, while the initial 438 GENFI cohort used T2-weighted images. Since white 439 matter hyperintensities do not reflect white matter 440 integrity, we performed DTI analysis in our sample, 441 where we did not find significant differences in our 442 groups after adjusting for age. Previous studies have 443 found white matter involvement in MAPT mutation 111 patients [22-25], particularly in the uncinate fascicu-445 lus [42, 44]. A recent study published in 2019 showed 446 a disproportional volume loss of the right temporal 447 lobe and more fractional anisotropy decline in the 448 uncinate fasciculus of MAPT carriers converting to 449 clinical FTD [45]. This study includes a follow-up 450 phase and compares converters, non-converters, and 451 non-carriers. Differences between groups were only 452 evident 2 years before symptom onset, while 4 years 453 before symptom onset these differences did not exist. 454 In our study we did not differentiate converters and 455 non-converters and we performed one cross-sectional 456 analysis, without controlling for time to onset. These 457 methodological disparities could explain the different 458 results regarding white matter integrity. 459

Studies performed in symptomatic subjects have been much more numerous, although once the disease is clinically noticeable, structural changes are generally widespread. Moreover, many of these studies are performed in sporadic forms of the disease, which may differ from genetic cases. However, neuropathological studies performed in sporadic FTD patients who died early in the course of their illness [5] show atrophy in some of the areas affected in our study: the frontal lobe, the medial temporal lobe (hippocampus, amygdala), and anterior cingulate gyrus. Some authors studying specifically neuroimaging changes in MAPT mutation patients [8, 40] report predominant gray matter loss in the temporal lobe, particularly the anterior and medial temporal lobe, with varying degrees of frontal and parietal lobe involvement in clinically diagnosed FTD. The orbitofrontal cortex, ventral insula, and anterior cingulate have also been found affected [7]. When subjects with mutations

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in the MAPT gene show a widespread pattern of 470 frontotemporal gray matter loss, the most severely 480 affected regions are the anteromedial temporal lobes, 481 suggesting that this may be the first area affected by 482 the disease [8]. MAPT patients tend to show sym-483 metric patterns of atrophy [46], with no differences 484 between left and right hemisphere when comparing 485 bilateral regions of interest. This is consistent with 486 our sample of preclinical mutation carriers where 487 most areas in both hemispheres were also equally 488 affected, suggesting that the disease begins and pro-489 gresses symmetrically. 490

Overall, the atrophy profile observed in MAPT 491 patients involves a ventral orbitofrontal-medial 492 temporal-ventral insula network [9]. Dysfunction 493 of this network has been associated with poor 494 performance on memory and naming, executive dys-495 function, and language deficits, widely recognized 496 in FTD [47]. Patients commonly develop semantic 497 impairment later in the disease, as well as prominent 498 episodic memory difficulties [9]. Other structures 499 affected in our sample (amygdala and cingulate cor-500 tex) belong to the rostral limbic system, which has 501 been suggested to underlie FTD symptoms [47]. 502 This system integrates limbic structures with output-503 related structures: the amygdala processes the value 504 of internal and external stimuli, represents that value 505 in the form of emotion to the brain and associates this 506 emotion to external stimuli. Moreover, the amygdala, 507 in close connection with the ventromedial prefrontal 508 and anterior cingulate cortices, contributes to other 509 higher order functions such as decision-making, the-510 ory of mind, and emotional processing [48, 49], while 511 the anterior section of the cingulate cortex detects 512 conflict within ongoing information processing and 513 integrates information from different structures of the 514 circuit [50]. 515

There is evidence that early changes in connectiv-516 ity could precede the occurrence of regional atrophy. 517 Some authors studied asymptomatic mutation car-518 riers using functional neuroimaging and reported 519 changes in connectivity, metabolic structure, and 520 blood flow without structural changes [18], suggest-521 ing that structural imaging changes may appear after 522 deficits in functional networks have been going on for 523 some time. Alberici et al. [51] reported significant 524 reductions of frontal lobe blood flow (dorsolateral 525 frontal cortex, frontal poles, and mesial frontal cor-526 tex) in an asymptomatic P301L mutation carrier using 527 SPECT, although these changes were not evident 528 in structural brain imaging. Dopper and colleagues 529 [41] reported frontal, posterior temporal, and pari-530

etal hypoperfusion in asymptomatic *MAPT* and *GRN* mutation carriers. Whitwell et al. [18] compared functional connectivity in *MAPT* mutation carriers, healthy controls, and bvFTD patients. Although there was no significant reduction of salience network connectivity in *MAPT* carriers, there was a suggestion of reduced connectivity in the anterior cingulate, one of the areas affected in our cohort's *MAPT* mutation carriers. The aforementioned studies suggest that once changes are noticeable using structural neuroimaging, deficits in functional brain networks may have been going on for some time.

This study was performed in a well-characterized and broadly phenotyped group of asymptomatic MAPT mutation carriers and familial matched controls. We would like to remark the strength of our findings: MAPT carriers and controls were recruited from the offspring generation of only five families, and were matched by age and family for analyses. Age, sex, and education were included in all analyses as covariates to further reduce potential confounds. The differences we found between carriers and non-carriers survived multiple comparisons and elastic/net analysis. Nevertheless, there were limitations to our study. First, our cohort is small when compared to previous studies. Second, although none of our carriers met diagnostic criteria for FTD, some of them received a CDR score of 0.5 for questionable symptoms and it is arguable if we can describe these patients as pre-symptomatic or if defining an MCI-FTD stage subgroup could be more appropriate. A CDR 0.5 allows a suspicion of early dementia, meaning that the subject shows consistent changes in cognitive function or functional impairments, even if they do not fulfill diagnostic criteria for FTD. It would be also desirable to know the expected time to onset for each subject, as it was reported in previous studies, and it would also help clarify the meaning of the CDR 0.5 subjects in this sample. As the specific underlying mutation may affect the pattern of atrophy [40], a subgroup analysis would have been desirable, but this was not possible due to the size of our sample. Specifically, patients carrying IVS10+16, IVS10+3, N279K, and S305N mutations show the most severe grey matter loss in the anterior temporal lobe, especially the medial structures. Patients with P301L or V337M mutations also show severe gray matter loss in the anterior temporal lobe, but unlike in our study, with a relative sparing of the medial temporal lobe and greater atrophy observed in more inferior and lateral temporal regions [40]. In addition, there is evidence that P301L and V337M FTD

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patients exhibit severe atrophy of the basal ganglia, 583 a finding that was observed in our cohort of MAPT 584 mutation carriers as a whole. These differences in 585 patterns of atrophy between MAPT mutations may 586 be secondary to their effects in the splicing of exon 587 10 and in the structural and functional properties of 588 the resulting tau protein [52]. Larger samples explor-580 ing how different mutations result in diverse atrophy 590 profiles are needed. 591

In conclusion, this study provides additional data 592 regarding early structural changes in a homogeneous 593 sample of preclinical MAPT mutation carriers, adding 594 to previous reports [20]. In our sample, atrophy was 595 detected in preclinical mutation carriers compared 596 to related non-carriers. Temporal lobe (left amyg-597 dala, left temporal pole), cingulate cortex (left rostral 598 anterior cingulate gyrus, right posterior cingulate), 599 and the lingual gyrus seem to be early targets of 600 the disease. Regarding white matter, we did not find 601 differences in white matter hyperintensities or DTI 602 analysis after adjusting for age. 603

Although this cross-sectional study offers valuable information, we continue to follow these patients in a longitudinal study design so we can assess atrophy rates across time. The degree to which FTD spreads between neighboring regions of the brain versus following a functional network comprised of spatially separated brain regions is still under investigation.

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# 622 SUPPLEMENTARY MATERIAL

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#### REFERENCES

- [1] Knopman DS, Roberts RO (2011) Estimating the number of persons with frontotemporal lobar degeneration in the US population. *J Mol Neurosci* **45**, 330-335.
- [2] Rohrer JD, Guerreiro R, Vandrovcova J, Uphill J, Reiman D, Beck J, Isaacs AM, Authier A, Ferrari R, Fox NC, Mackenzie IR, Warren JD, de Silva R, Holton J, Revesz T, Hardy J, Mead S, Rossor MN (2009) The heritability and genetics of frontotemporal lobar degeneration. *Neurology* **73**, 1451-1456.
- [3] Rohrer JD, Warren JD (2011) Phenotypic signatures of genetic frontotemporal dementia. *Curr Opin Neurol* 24, 542-549.
- [4] Whitwell JL, Weigand SD, Boeve BF, Senjem ML, Gunter JL, DeJesus-Hernandez M, Rutherford NJ, Baker M, Knopman DS, Wszolek ZK, Parisi JE, Dickson DW, Petersen RC, Rademakers R, Jack CR Jr, Josephs KA (2012) Neuroimaging signatures of frontotemporal dementia genetics: C90RF72, tau, progranulin and sporadics. *Brain* 135, 794-806.
- [5] Kril JJ, Halliday GM (2004) Clinicopathological staging of frontotemporal dementia severity: Correlation with regional atrophy. *Dement Geriatr Cogn Disord* 17, 311-315.
- [6] Mahoney CJ, Beck J, Rohrer JD, Lashley T, Mok K, Shakespeare T, Yeatman T, Warrington EK, Schott JM, Fox NC, Rossor MN, Hardy J, Collinge J, Revesz T, Mead S, Warren JD (2012) Frontotemporal dementia with the C9ORF72 hexanucleotide repeat expansion: Clinical, neuroanatomical and neuropathological features. *Brain* 135, 736-750.
- [7] Rohrer JD, Ridgway GR, Modat M, Ourselin S, Mead S, Fox NC, Rossor MN, Warren JD (2010) Distinct profiles of brain atrophy in frontotemporal lobar degeneration caused by progranulin and tau mutations. *Neuroimage* 53, 1070-1076.
- [8] Whitwell JL, Jack CR Jr, Boeve BF, Senjem ML, Baker M, Rademakers R, Ivnik RJ, Knopman DS, Wszolek ZK, Petersen RC, Josephs KA (2009) Voxel-based morphometry patterns of atrophy in FTLD with mutations in MAPT or PGRN. *Neurology* **72**, 813-820.
- [9] Seeley WW, Crawford R, Rascovsky K, Kramer JH, Weiner M, Miller BL, Gorno-Tempini ML (2008) Frontal paralimbic network atrophy in very mild behavioral variant frontotemporal dementia. *Arch Neurol* 65, 249-255.
- [10] Van Swieten J, Spillantini MG (2007) Hereditary frontotemporal dementia caused by Tau gene mutations. *Brain Pathol* 17, 63-73.
- [11] Ghetti B, Oblak AL, Boevet BF, Johnson KA, Dickerson BC, Goedert M (2015) Invited review: Frontotemporal dementia caused by *microtubule-associated protein tau* (*MAPT*) mutations: A chameleon for neuropathology and neuroimaging. *Neuropathol Appl Neurobiol* **41**, 24-46.
- [12] Cheran G, Silverman H, Manoochehri M, Goldman J, Lee S, Wu L, Cines S, Fallon E, Kelly BD, Olszewska DA, Heidebrink J, Shair S, Campbell S, Paulson H, Lynch T, Cosentino S, Huey ED (2017) Psychiatric symptoms in preclinical behavioural-variant frontotemporal dementia in *MAPT* mutation carriers. *J Neurol Neurosurg Psychiatry* 89, 449-455.
- [13] Eskildsen SF, Østergaard LR, Rodell AB, Østergaard L, Nielsen JE, Isaacs AM, Johannsen P (2009) Cortical volumes and atrophy rates in FTD-3 CHMP2B mutation carriers and related non-carriers. *Neuroimage* 45, 713-721.

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- [14] Rohrer JD, Ahsan RL, Isaacs AM, Nielsen JE, Ostergaard
  L, Scahill R, Warren JD, Rossor MN, Fox NC, Johannsen
  P; FReJA consortium (2009) Presymptomatic generalized brain atrophy in frontotemporal dementia caused by
  CHMP2B mutation. *Dement Geriatr Cogn Disord* 27, 182-186.
- [15] Lunau L, Mouridsen K, Rodell A, Ostergaard L, Nielsen JE,
   Isaacs A, Johannsen P; FReJA Consortium (2012) Presymp tomatic cerebral blood flow changes in CHMP2B mutation
   carriers of familial frontotemporal dementia (FTD-3), mea sured with MRI. *BMJ Open* 2, e000368.
- [16] Walhout R, Schmidt R, Westeneng HJ, Verstraete E, Seelen M, van Rheenen W, de Reus MA, van Es MA, Hendrikse J, Veldink JH, van den Heuvel MP, van den Berg LH (2015) Brain morphologic changes in asymptomatic C90rf72 repeat expansion carriers. *Neurology* 85, 1780-1788.
- [17] Pievani M, Paternicò D, Benussi L, Binetti G, Orlandini
   A, Cobelli M, Magnaldi S, Ghidoni R, Frisoni GB (2014)
   Pattern of structural and functional brain abnormalities
   in asymptomatic granulin mutation carriers. *Alzheimers Dement* 10, S354-S63 e1.
- [18] Whitwell JL, Josephs KA, Avula R, Tosakulwong N,
  Weigand SD, Senjem ML, Vemuri P, Jones DT, Gunter
  JL, Baker M, Wszolek ZK, Knopman DS, Rademakers
  R, Petersen RC, Boeve BF, Jack CR Jr (2011) Altered
  functional connectivity in asymptomatic MAPT subjects:
  A comparison to bvFTD. *Neurology* 77, 866-874.
- [19] Rohrer JD, Nicholas JM, Cash DM, van Swieten J, Dopper 714 E, Jiskoot L, van Minkelen R, Rombouts SA, Cardoso MJ, 715 Clegg S, Espak M, Mead S, Thomas DL, De Vita E, Masel-716 717 lis M, Black SE, Freedman M, Keren R, MacIntosh BJ, 718 Rogaeva E, Tang-Wai D, Tartaglia MC, Laforce R Jr, Tagliavini F, Tiraboschi P, Redaelli V, Prioni S, Grisoli M, Borroni 719 B, Padovani A, Galimberti D, Scarpini E, Arighi A, Fuma-720 galli G, Rowe JB, Coyle-Gilchrist I, Graff C, Fallström M, 721 Jelic V, Ståhlbom AK, Andersson C, Thonberg H, Lilius L, 722 Frisoni GB, Pievani M, Bocchetta M, Benussi L, Ghidoni R, 723 724 Finger E, Sorbi S, Nacmias B, Lombardi G, Polito C, Warren JD, Ourselin S, Fox NC, Rossor MN, Binetti G (2015) 725 Presymptomatic cognitive and neuroanatomical changes in 726 genetic frontotemporal dementia in the Genetic Frontotem-727 poral dementia Initiative (GENFI) study: A cross-sectional 728 analysis. Lancet Neurol 14, 253-262. 729
- 730 [20] Cash DM, Bocchetta M, Thomas DL, Dick KM, van Swieten JC, Borroni B, Galimberti D, Masellis M, Tartaglia MC, 731 Rowe JB, Graff C, Tagliavini F, Frisoni GB, Laforce R Jr, 732 Finger E, de Mendonça A, Sorbi S, Rossor MN, Ourselin S, 733 Rohrer JD (2018) Genetic FTD Initiative, GENFI. Patterns 734 of gray matter atrophy in genetic frontotemporal demen-735 tia: Results from the GENFI study. Neurobiol Aging 62, 736 191-196 737
- [21] Miyoshi M, Shinotoh H, Wszolek ZK, Strongosky AJ, Shimada H, Arakawa R, Higuchi M, Ikoma Y, Yasuno F,
  Fukushi K, Irie T, Ito H, Suhara T (2010) *In vivo* detection of neuropathologic changes in presymptomatic MAPT mutation carriers: A PET and MRI study. *Parkinsonism Relat Disord* 16, 404-408.
- [22] Mahoney CJ, Ridgway GR, Malone IB, Downey LE, Beck
  J, Kinnunen KM, Schmitz N, Golden HL, Rohrer JD, Schott
  JM, Rossor MN, Ourselin S, Mead S, Fox NC, Warren JD
  (2014) Profiles of white matter tract pathology in frontotemporal dementia. *Hum Brain Mapp* 35, 4163-4179.
- [23] Tacik P, Sanchez-Contreras M, DeTure M, Murray ME,
   Rademakers R, Ross OA, Wszolek ZK, Parisi JE, Knopman

DS, Petersen RC, Dickson DW (2017) Clinicopathologic heterogeneity in frontotemporal dementia and parkinsonism linked to chromosome 17 (FTDP-17) due to microtubuleassociated protein tau (MAPT) p.P301L mutation, including a patient with globular glial tauopathy. *Neuropathol Appl Neurobiol* **43**, 200-214.

- [24] Caroppo P, Le Ber I, Camuzat A, Clot F, Naccache L, Lamari F, De Septenville A, Bertrand A, Belliard S, Hannequin D, Colliot O, Brice A (2014) Extensive white matter involvement in patients with frontotemporal lobar degeneration. *JAMA Neurol* **71**, 1562.
- [25] Paternicò D, Premi E, Gazzina S, Cosseddu M, Alberici A, Archetti S, Cotelli MS, Micheli A, Turla M, Gasparotti R, Padovani A, Borroni B (2016) White matter hyperintensities characterize monogenic frontotemporal dementia with granulin mutations. *Neurobiol Aging* 38, 176-180.
- [26] Sudre CH, Bocchetta M, Cash D, Thomas DL, Woollacott I, Dick KM, van Swieten J, Borroni B, Galimberti D, Masellis M, Tartaglia MC, Rowe JB, Graff C, Tagliavini F, Frisoni G, Laforce R Jr, Finger E, de Mendonça A, Sorbi S, Ourselin S, Cardoso MJ, Rohrer JD, Genetic FTD Initiative, GENFI (2017) White matter hyperintensities are seen only in GRN mutation carriers in the GENFI cohort. *Neuroimage Clin* 15, 171-180.
- [27] Petersen ET, Mouridsen K, Golay X; all named co-authors of the QUASAR test-retest study (2010) The QUASAR reproducibility study, Part II: Results from a multi-center Arterial Spin Labeling test-retest study. *Neuroimage* 49, 104-113.
- [28] Walhovd KB, Westlye LT, Amlien I, Espeseth T, Reinvang I, Raz N, Agartz I, Salat DH, Greve DN, Fischl B, Dale AM, Fjell AM (2011) Consistent neuroanatomical age-related volume differences across multiple samples. *Neurobiol Aging* 32, 916-932.
- [29] Whitwell JL, Jack CRJ (2005) Comparisons between Alzheimer disease, frontotemporal lobar degeneration, and normal aging with brain mapping. *Top Magn Reson Imaging* 16, 409-425.
- [30] Smith S.M, Jenkinson M, Woolrich MW, Beckmann CF, Behrens TEJ, Jihansen-Berg H, Bannister PR, De Luca M, Drobnjak I, Flitney Dem Niazy R, Saunders J, Vickers J, Zhang Y, De Stefano N, Brady JM, Matthews PM (2004) Advances in functional and structural MR image analysis and implementation as FSL. *Neuroimage* 23, 208-219.
- [31] Smith M, Jenkinson M, Johansen-Berg H, Rueckert D, Nichols TE, Mackay CE, Watkins KE, Ciccarelli O, Cader MZ, Matthews PM, Behrens TEJ (2006) Tract-based spatial statistics: Voxelwise analysis of multi-subject diffusion data. *Neuroimage* 31, 1487 1505.
- [32] Andersson JLR, Jenkinson M, Smith S (2007) Nonlinear optimisation. FMRIB technical report TR07JA1. http://www.fmrib.ox.ac.uk/analysis/techrep.
- [33] Andersson JLR, Jenkinson M, Smith S (2007) Nonlinear registration, aka Spatial normalisation FMRIB technical report TR07JA2. http://www.fmrib.ox.ac.uk/ analysis/techrep.
- [34] Mori S, Wakana S, van Zijl PCM, Nagae-Poetscher LM (2005) Atlas of Human White Matter. Elsevier, Amsterdam.
- [35] Friedman J, Hastie T, Tibshirani R (2010) Regularization paths for generalized linear models via coordinate descent. J Stat Softw 2010 33, 1-22. Available at: http://www.jstatsoft.org/v33/i01/.
- [36] Robin X, Turck N, Hainard A, Tiberti N, Lisacek F, Sanchez JC, Müller M (2011) pROC: An open-source package for R

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and S+ to analyze and compare ROC curves. BMC Bioinformatics 12, 77.

- Benjamini Y, Hochberg Y (1995) Controlling the false dis-818 [37] covery rate: A practical and powerful approach to multiple 819 820 testing. J R Stat Soc Series B 57, 289-300.
- [38] Hui Z, Hastie T (2005) Regularization and variable selection 821 via the elastic net. J R Statist Soc B 67(Pt 2), 301-320. 822
- [39] Knopman DS, Kramer JH, Boeve BF, Caselli RJ, Graff-823 824 Radford NR, Mendez MF, Miller BL, Mercaldo N (2008) Development of methodology for conducting clinical trials 825 in frontotemporal lobar degeneration. Brain 131, 2957-826 2968 827
- Whitwell JL, Jack CR Jr, Boeve BF, Senjem ML, Baker [40] 828 M, Ivnik RJ, Knopman DS, Wszolek ZK, Petersen RC, 829 Rademakers R, Josephs KA (2009) Atrophy patterns in 830 IVS10+16, IVS10+3, N279K, S305N, P301L, and V337M 831 MAPT mutations. Neurology 73, 1058-1065. 832
- [41] Dopper EG, Chalos V, Ghariq E, den Heijer T, Hafkemei-833 jer A, Jiskoot LC, de Koning I, Seelaar H, van Minkelen 834 R, van Osch MJ, Rombouts SA, van Swieten JC (2016) 835 836 Cerebral blood flow in presymptomatic MAPT and GRN 837 mutation carriers: A longitudinal arterial spin labeling study. Neuroimage Clin 12, 460-465. 838
- Panman JL, Jiskoot LC, Bouts MJRJ, Meeter LHH, van der 839 [42] Ende EL, Poos JM, Feis RA, Kievit AJA, van Mikelen R, 840 Dopper EGP, Rombouts SARB, van Swieten JC, Papma JM 841 842 (2019) Gray and white matter changes in presymptomatic genetic frontotemporal dementia: A longitudinal MRI study. 843 Neurobiol Aging 76, 115-124. 844
- 845 [43] Forman MS (2004) Genotype-phenotype correlations in FTDP-17: Does form follow function? Exp Neurol 187, 846 847 229-234.
- 848 [44] Jiskoot LC, Bocchetta M, Nicholas JM, Cash DM, Thomas D, Modat M, Ourselin S, Rombouts SARB, Dopper EGP, 849 Meeter LH, Panman JL, van Minkelen R, van der Ende EL, 850 Donker Kaat L, Pijnenburg YAL, Borroni B, Galimberti D, 851 Masellis M, Tartaglia MC, Rowe J, Graff C, Tagliavini F, 852 Frisoni GB, Laforce R Jr, Finger E, de Mendonça A, Sorbi 853 S, on behalf of the Genetic Frontotemporal dementia Initia-

tive (GENFI), Papma JM, van Swieten JC, Rohrer JD (2018) Presymptomatic white matter integrity loss in familial frontotemporal dementia in the GENFI cohort: A cross-sectional diffusion tensor imaging study. Ann Clin Transl Neurol 5, 1025-1036.

- [45] Jiskoot LC, Panman JL, Meeter LH, Dopper EGP, Donker Kaat L, Franzen S, van der Ende EL, van Minkelen R, Rombouts SARB, Papma JM, van Swieten JC (2019) Longitudinal multimodal MRI as prognostic and diagnostic biomarker in presymptomatic familial frontotemporal dementia. Brain 142, 193-208.
- [46] Van Deerlin VM, Wood EM, Moore P, Yuan W, Forman MS, Clark CM, Neumann M, Kwong LK, Trojanowski JQ, Lee VM, Grossman M (2007) Clinical, genetic, and pathologic characteristics of patients with frontotemporal dementia and progranulin mutations. Arch Neurol 64, 1148-1153.
- [47] Boccardi M, Sabattoli F, Laakso MP, Testa C, Rossi R, Beltramello A, Soininen H, Frisoni GB (2005) Frontotemporal dementia as a neural system disease. Neurobiol Aging 26, 37-44.
- [48] Cardinal RN, Parkinson JA, Hall J, Everitt, BJ (2002) Emotion and motivation: The role of the amygdala, ventral striatum, and prefrontal cortex. Neurosci Biobehav Rev 26, 321-352
- Stone VE, Baron-Cohen S, Calder A, Keane J, Young A [49] (2003) Acquired theory of mind impairments in individuals with bilateral amygdala lesions. Neuropsychologia 41, 209-220
- Devinsky O, Morrell MJ, Vogt BA (1995) Contributions of [50] anterior cingulate cortex to behavior. Brain 118, 279-306.
- [51] Alberici A, Gobbo C, Panzacchi A, Nicosia F, Ghidoni R, Benussi L, Hock C, Papassotiropoulos A, Liberini P, Growdon JH, Frisoni GB, Villa A, Zanetti O, Cappa S, Fazio F, Binetti G (2004) Frontotemporal dementia: Impact of P301L tau mutation on a healthy carrier. J Neurol, Neurosurg Psychiatry 75, 1607-1610.

Goedert M, Jakes R (2005) Mutations causing neurodegen-

erative tauopathies. Biochim Biophys Acta 1739, 240-250.

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